



BIOORGANIC CHEMISTRY

www.elsevier.com/locate/bioorg

Bioorganic Chemistry 35 (2007) 25-34

Synthesis of pyrrolo[2,3-d]pyrimidine nucleoside derivatives as potential anti-HCV agents

Chamakura V.N.S. Varaprasad *, Kanda S. Ramasamy, Jean-Luc Girardet, Esmir Gunic, Vicky Lai, Weidong Zhong, Haoyun An ¹, Zhi Hong

Drug Discovery, Valeant Pharmaceuticals Research and Development, 3300 Hyland Avenue, Costa Mesa, CA 92626, USA

Received 15 June 2006 Available online 1 September 2006

Abstract

Several Toyocamycin (4) analogues were examined for their ability to inhibit HCV RNA in a replicon assay. Among the compounds examined 4-methylthio (18) and 5-carboxamide oxime derivatives (23 and 27) of Toyocamycin were found to have good activity and selectivity. © 2006 Elsevier Inc. All rights reserved.

Keywords: HCV NS5B inhibitors; Toyocamycin; Pyrrolo[2,3-d]pyrimidine nucleosides

1. Introduction

Hepatitis C virus (HCV) infection is highly persistent and lacks an effective and well tolerated antiviral therapy. This has spurred intensive efforts to discover novel anti-HCV agents. The RNA-dependent RNA polymerase (RdRp), NS5B of HCV, constitutes a valid target for drug discovery as it is the centerpiece for viral replication. Compared to the host RNA and DNA polymerases, NS5B RdRp has distinct sub cellular localization at the interface of the endoplasmic reticulum membrane and cytoplasm, a novel catalytic

E-mail address: vchamakura@gmail.com (C.V.N.S. Varaprasad).

^{*} Corresponding author. Fax: +1 714 641 7233.

¹ Present address: ACEA Biosciences, Inc., 11585 Sorrento Valley Rd., Ste.103, San Diego, CA 92121, USA.

HO OH OH
$$R^5$$
 $R^5 = H; R^7 = H$ $R^5 = H; R^7 = ribofuranosyl$ $R^5 = CN; R^7 = ribofuranosyl$

Fig. 1. Adenosine lead derivative (1) and Toyocamycin (4) analogues.

mechanism and many unique structural features, all of which make it an attractive target for developing effective anti-HCV therapeutics [1].

Currently, combinations of pegylated interferons and ribavirin are the leading therapy [2] for hepatitis C virus. However, even these combinations have limited efficacy. Many classes of nucleoside and non-nucleoside inhibitors of NS5B RdRp have been identified and appeared in literature and patent applications in pursuit of a better treatment for HCV.

Our continued interest [3] to develop a promising treatment for HCV infection has led us to screen our nucleoside library against HCV polymerase in a replicon assay. This effort identified N⁶,N⁶-dimethyl adenosine (1, Fig. 1) as the lead compound (EC₅₀: 7 μM, CC₅₀: 300 μM). Whereas Toyocamycin [4] (4) has long been, along with its congener nucleosides Tubercidin (3) and Sangivamycin (5), studied for its antiviral (human cytomegalovirus, [HCMV]) and antitumor potencies. Accordingly, Toyocamycin analogues comprising nucleoside [5] and the related heterocycle (2), pyrrolo[2,3-d]pyrimidine [6], modifications were examined against HCMV. The nucleoside modifications included both of sugar and aglycone moieties while pyrrolo[2,3-d]pyrimidine 2 heterocycle modifications involved C5 and N7 positions. As part of SAR studies in order to improve the activity of lead adenosine derivative 1 several pyrrolo[2,3-d]pyrimidine nucleoside analogues related to Toyocamycin 4 were synthesized and screened for their potential in the inhibition of HCV polymerase in replicon assay [7]. In this report, we disclose results of our study.

2. Materials and methods

2.1. General

 1 H NMR spectra were obtained using Varian Gemini 300 MHz instrument. Majority of reagents were procured from Aldrich Chemical Company, MI, USA and used as received. All anhydrous reactions were carried out under Argon atmosphere. The reactions were monitored either by thin layer chromatography (TLC), using TLC sheets coated with silica gel 60 F₂₅₄ (Merck) and UV light visualization, or by LC-MS (waters) Micromass). The products were purified by flash chromatography on Merck silica gel 60 (230–400 mesh ASTM). The replicon assay, in presence of the inhibitors, was carried out as described in the literature [7].

2.2. 3,4-Dihydro-4-oxo-7-(β-D-ribofuranosyl)pyrrolo[2,3-d]pyrimidine-5-carbonitrile (6)

To a solution of Toyocamycin **4** (7.275 g, 25 mmol) in a mixture of distilled water (200 ml) and glacial acetic acid (50 ml) solid NaNO₂ (17.25 g, 250 mmol) was slowly added at 50 °C. The reaction mixture was heated at 70 °C for 1 h. The volatiles were evaporated under reduced pressure to dryness and the residue was extracted with hot acetone (4× 100 ml). The combined acetone extracts were evaporated and the residue was purified over flash silica gel chromatography to obtain the title product (6.13 g, 84%). ¹H NMR (CD₃OD): δ 8.18 (s, 1H), 8.00 (s, 1H), 6.14 (d, J = 5.4 Hz, 1H), 4.43 (t, J = 5.4 Hz, 1H), 4.27 (t, J = 3.9, 5.4 Hz, 1H), 4.10 (dd, J = 3.0, 6.6 Hz, 1H), 3.80 (ddd, J = 2.7, 12.0 and 32 Hz, 2H). ES-MS (m/z+1) $^+$: 293.

2.3. 3,4-Dihydro-4-oxo-7-(2,3,5-tri-O-acetyl- β -D-ribofuranosyl)pyrrolo[2,3-d]pyrimidine-5-carbonitrile (7)

To a solution of 3,4-dihydro-4-oxo-7-(β-D-ribofuranosyl)pyrrolo[2,3-d]pyrimidine-5-carbonitrile **6** (5.84 g, 20 mmol) in pyridine (50 ml) acetic anhydride (8.46 ml, 90 mmol) was added and stirred at room temperature for 16 h. The volatiles evaporated and the residue was dissolved in a mixture of CH₂Cl₂ (200 ml) and water (200 ml). The organic layer was separated and washed with water (200 ml), aq. HCl (1 N, 2× 200 ml), and water (200 ml), dried (MgSO₄) and evaporated over a rotary evaporator. The residue was purified over flash silica gel chromatography (CH₂Cl₂/MeOH using as eluants with increasing polarity of the mixture) to obtain the titled product as a colorless solid (6.94 g, 83%). ¹H NMR (CDCl₃): δ 12.69 (br s, D₂O exchangeable, 1H), 8.13 (s, 1H), 7.69 (s, 1H), 6.31 (d, J = 5.4 Hz, 1H), 5.62 (t, J = 5.4 Hz, 1H), 5.47 (dd, J = 4.2, 5.4 Hz, 1H), 4.45 (m, 1H), 4.40 (d, J = 4.2 Hz, 2H), 2.18 (s, 3H), 2.15 (s, 3H), 2.09 (s, 3H). ES-MS (m/z+1)⁺: 419.

2.4. 4-Chloro-7-(2,3,5-tri-O-acetyl- β -D-ribofuranosyl)pyrrolo[2,3-d]pyrimidine-5-carbonitrile ($\pmb{8}$)

3,4-Dihydro-4-oxo-7-(2,3,5-tri-O-acetyl- β -D-ribofuranosyl)pyrrolo[2,3-d]pyrimidine-5-carbonitrile, 7 (4.18 g, 10 mmol) was dissolved in phosphorous oxychloride (25 ml) and refluxed for 1 h. The reaction mixture was cooled to room temperature and the volatiles evaporated. The residue was suspended in ice cold water and stirred vigorously. The solids separated were filtered and dried (P_2O_5) under reduced pressure to obtain the titled crude product in a quantitative yield. The hygroscopic solid was used in the next reactions without further purification. ES-MS $(m/z+1)^+$: 437.

2.5. The following procedure for the synthesis of 4-methylamino-7-(β -D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine-5-carbonitrile 10 exemplifies the general method for the synthesis of mono N^4 -alkyl Toyocamycin derivatives 10–15. The rest of the compounds in the series were characterized by LC-MS analysis

4-Chloro-7-(2,3,5-tri-O-acetyl-β-D-ribofuranosyl)pyrrolo[2,3-d]pyrimidine-5-carbonitri-le **8** (0.3 mmol) was treated with a solution of methylamine (3 ml, 2 M) in THF was added and the reaction flask was closed tightly. The reaction mixture stirred at room temperature for 16 h. The volatiles were evaporated and the residue was purified over flash silica gel

chromatography using CH₂Cl₂ and MeOH as eluants with increasing polarity to obtain 4-methylamino-7-(β -D-ribofuranosyl)pyrrolo[2,3-d]pyrimidine-5-carbonitrile **10** (63%). ¹H NMR (CD₃OD): δ 8.13 (s, 1H), 8.02 (s, 1H), 6.00 (d, J = 6.3 Hz, 1H), 4.62 (dd, J = 5.4, 6.3 Hz, 1H), 4.27 (dd, J = 3.0, 5.4 Hz, 1H), 4.13 (dd, J = 3.0, 6.0 Hz, 1H), 3.80 (ddd, J = 2.4, 12.3 and 33.2 Hz, 2H), 3.05 (s, 3H). ES-MS (m/z+1)⁺: 306.

2.6. 4-Chloro-7-(β-D-ribofuranosyl)pyrrolo[2,3-d]pyrimidine-5-carbonitrile (9)

4-Chloro-7-(2,3,5-tri-*O*-acetyl-β-D-ribofuranosyl)pyrrolo[2,3-*d*]pyrimidine-5-carbonitrile **8** (1.9 g, 4.36 mmol) was treated with a cold (-20 °C) solution of saturated methanolic ammonia and the reaction mixture kept at 5 °C for 2.5 h with occasional shaking. The volatiles evaporated and the residue was purified over flash silica gel chromatography using CH₂Cl₂ and MeOH mixture with increasing polarity to obtain the title product (1.1 g, 81.5%). ¹H NMR (CD₃OD): δ 8.16 (s, 1H), 8.01 (s, 1H), 6.12 (d, J = 5.4 Hz, 1H), 4.40 (t, J = 5.4 Hz, 1H), 4.27 (t, J = 3.9, 5.4 Hz, 1H), 4.09 (dd, J = 3.0, 6.6 Hz, 1H), 3.80 (ddd, J = 2.7, 12.0 and 32 Hz, 2H). ES-MS (m/z+1)⁺: 312.

2.7. 4-Dimethylamino-7-(β-D-ribofuranosyl)pyrrolo[2,3-d]pyrimidine-5-carbonitrile (16)

To a solution of 4-chloro-7-(β-D-ribofuranosyl)pyrrolo[2,3-d]pyrimidine-5-carbonitrile **9** (0.3 g, 0.96 mmol) in EtOH (10 ml) a solution of dimethylamine (10 ml, 2 M) in THF was added and the reaction mixture was refluxed for 1 h. The volatiles evaporated and the residue was purified by flash silica gel chromatography using CH₂Cl₂/MeOH mixture with increasing polarity to obtain the titled product (0.2 g, 65.15%). ¹H NMR (CD₃OD): δ 8.34 (s, 1H), 8.24 (s, 1H), 6.13 (d, J = 5.1 Hz, 1H), 4.52 (t, J = 5.1 Hz, 1H), 4.27 (dd, J = 3.9, 5.1 Hz, 1H), 4.12 (dd, J = 2.7, 6.6 Hz, 1H), 3.82 (ddd, J = 2.4, 12.3 and 37.5 Hz, 2H), 3.39 (s, 6H). ES-MS (m/z+1)⁺: 320.

2.8. 4-O-Methyl-7-(β-D-ribofuranosyl)pyrrolo[2,3-d]pyrimidine-5-carbonitrile (17)

A solution of 4-chloro-7-(β -D-ribofuranosyl)pyrrolo[2,3-d]pyrimidine-5-carbonitrile **9** (0.310 g, 1 mmol) and sodium methoxide (0.054 g, 5 mmol) in methyl alcohol (10 ml) was refluxed for 16 h. The reaction mixture was cooled to room temperature and volatiles evaporated. The residue was purified over flash silica gel chromatography (CH₂Cl₂/MeOH mixture as eluant) to obtain the titled product (0.16 g, 52%) as a white solid. ¹H NMR (CD₃OD): δ 8.77 (s, 1H), 8.58 (s, 1H), 6.20 (d, J = 5.4 Hz, 1H), 4.53 (t, J = 5.4 Hz, 1H), 4.22 (dd, J = 3.9, 4.8 Hz, 1H), 4.13 (dd, J = 3.0, 7.2 Hz, 1H), 3.80 (ddd, J = 2.7, 12.3 and 34.2 Hz, 2H), 4.02 (s, 3H). ES-MS (m/z+1)⁺: 307.

2.9. 4-Methylthio-7-(β-p-ribofuranosyl)pyrrolo[2,3-d]pyrimidine-5-carbonitrile (18)

A suspension of 4-chloro-7-(β -D-ribofuranosyl)pyrrolo[2,3-d]pyrimidine-5-carbonitrile 9 (0.310 g, 1 mmol) and sodium methanethiolate (0.105 g, 1.5 mmol) in isopropyl alcohol (10 ml) was refluxed for 1 h. The reaction mixture cooled to room temperature and volatiles evaporated. The residue was purified over flash silica gel chromatography (CH₂Cl₂/MeOH mixture as eluant) to obtain the titled product (0.23 g, 72%) as a white solid. ¹H NMR (CD₃OD): δ 8.67 (s, 1H), 8.48 (s, 1H), 6.23 (d, J = 5.4 Hz, 1H), 4.51 (t,

J = 5.4 Hz, 1H), 4.29 (dd, J = 3.9, 4.8 Hz, 1H), 4.12 (dd, J = 3.0, 7.2 Hz, 1H), 3.82 (ddd, J = 2.7, 12.3 and 34.2 Hz, 2H), 2.72 (s, 3H). ES-MS $(m/z+1)^+$: 322.

2.10. 4-Methylamino-7- $(\beta$ -D-ribofuranosyl) pyrrolo[2,3-d]pyrimidine-5-carboxamidoxime (23)

To a solution of 4-methylamino-7-(β-D-ribofuranosyl)pyrrolo[2,3-d]pyrimidine-5-carbonitrile **10** (0.305 g, 1 mmol) in EtOH (10 ml) an aqueous solution of hydroxylamine (0.5 ml, 50% w/w) was added and the reaction mixture refluxed for 16 h. The volatiles were evaporated and the residue was recrystallized from minimum amount of methanol to obtain the title product (0.21 g, 62%) as a white solid. ¹H NMR (CD₃OD) δ : 8.07 (s, 1H), 7.68 (s, 1H), 5.97 (d, J = 6.6 Hz, 1H), 4.61 (dd, J = 5.4, 6.6 Hz, 1H), 4.26 (dd, J = 2.7, 5.1 Hz, 1H), 4.10 (dd, J = 3.0, 5.7 Hz, 1H), 3.79 (ddd, J = 34.8, 12.3 and 2.7 Hz, 2H), 3.02 (s, 3H). ES-MS (m/z+1) $^+$: 339.

2.11. 4-Methylamino-7-(β-D-ribofuranosyl) pyrrolo[2,3-d]pyrimidine-5-carboxamidine (24)

To a solution of 4-methylamino-7-(β-D-ribofuranosyl)pyrrolo[2,3-d]pyrimidine-5-carboxamidoxime **23** (0.14 g, 0.41 mmol) in a mixture of ethanol and water (10 ml, 1:1) ammonium chloride (0.022 g, 0.41 mmol) and thoroughly washed Raney nickel (0.5 ml) in water was successively added. The reaction mixture was shaken well under hydrogen atmosphere (50 psi) at room temperature for 16 h, filtered over celite and the filtrate was evaporated to obtain the titled product (0.1 g, 67%) as an hydrochloride salt. ¹H NMR (CD₃OD) δ: 8.28 (s, 1H), 8.14 (s, 1H), 6.10 (d, J = 5.7 Hz, 1H), 4.59 (t, J = 5.4 Hz, 1H), 4.29 (dd, J = 4.8, 3.3 Hz, 1H), 4.14 (dd, J = 6.3, 3.0 Hz, 1H), 3.79 (ddd, J = 32.4, 12.3 and 2.4 Hz, 2H), 3.10 (s, 3H). ES-MS $(m/z+1)^+$: 323.

2.12. 4-Dimethylamino-7-(β-D-ribofuranosyl)pyrrolo[2,3-d]pyrimidine-5-carboxamidoxime (25)

To a solution of 4-dimethylamino-7-(β -D-ribofuranosyl)pyrrolo[2,3-d]pyrimidine-5-carbonitrile, **16** (0.1 g, 0.31 mmol) in EtOH (10 ml) an aqueous solution of hydroxylamine (0.5 ml, 50% w/w) was added and the reaction mixture refluxed for 16 h. The volatiles were evaporated and the residue was purified over flash silica gel chromatography (CH₂Cl₂/MeOH mixture as eluant) to obtain the title product (0.06 g, 54.5%) as a white solid. ¹H NMR (CD₃OD) δ : 8.14 (s, 1H), 7.57 (s, 1H), 6.06 (d, J = 6.0 Hz, 1H), 4.58 (t, J = 5.7 Hz, 1H), 4.27 (dd, J = 5.4, 3.0 Hz, 1H), 4.09 (dd, J = 6.0, 2.7 Hz, 1H), 3.77 (ddd, J = 33.3, 12.3 and 2.7 Hz, 2H), 3.17 (s, 6H). ES-MS (m/z+1) $^+$: 353.

2.13. 4-Dimethylamino-7-(β-D-ribofuranosyl)pyrrolo[2,3-d]pyrimidine-5-carboxamidine (26)

To a solution of 4-dimethylamino-7-(β-D-ribofuranosyl)pyrrolo[2,3-d]pyrimidine-5-carboxamidoxime **25** (0.1 g, 0.28 mmol) in a mixture of ethanol and water (5 ml, 1:1) ammonium chloride (0.015 g, 0.28 mmol) and thoroughly washed Raney nickel (0.5 ml) in water was successively added. The reaction mixture was shaken well under hydrogen atmosphere (50 psi) at room temperature for 16 h. The reaction mixture was filtered over

celite and the filtrate evaporated to obtain the titled product (0.05 g, 49.5%) as an hydrochloride salt. ¹H NMR (CD₃OD) δ : 8.29 (s, 1H), 8.23 (s, 1H), 6.16 (d, J = 6.0 Hz, 1H), 4.58 (t, J = 5.4 Hz, 1H), 4.30 (dd, J = 4.5, 3.9 Hz, 1H), 4.13 (br d, J = 2.7 Hz, 1H), 3.82 (ddd, J = 29.1, 12.6 and 2.7 Hz, 2H), 3.14 (s, 6H). ES-MS (m/z+1)⁺: 337.

2.14. 4-Methythio-7-(β-D-ribofuranosyl)pyrrolo[2,3-d]pyrimidine-5-carboxamidoxime, (27)

To a solution of 4-methylthio-7-(β-D-ribofuranosyl)pyrrolo[2,3-d]pyrimidine-5-carbonitrile, **18** (0.2 g, 0.62 mmol) in EtOH (10 ml) an aqueous solution of hydroxylamine (0.5 ml, 50% w/w) was added and the reaction mixture refluxed for 16 h. The volatiles were evaporated and the residue was purified over flash silica gel chromatography (CH₂Cl₂/MeOH mixture as eluant) to obtain the product (0.21 g, 72.7%) as a white solid. ¹H NMR (CD₃OD) δ: 8.57 (s, 1H), 7.81(s, 1H), 6.21 (d, J = 5.4 Hz, 1H), 4.56 (t, J = 6.0 Hz, 1H), 4.29 (br m, 1H), 4.10 (br m,1H), 3.79 (br m, 2H), 2.62 (s, 3H). ES-MS $(m/z+1)^+$: 356.

Scheme 1. Reagents: (i) NaNO2, water, glacial AcOH; (ii) Ac2O, Py.; (iii) POCl3.

3. Results and discussion

The present study involved modifications of exocyclic amino group and cyano group, at C4 and C5 positions, respectively, of Toyocamycin (4). To explore the various substitutions tolerated on exocyclic amino group several N^4 -alkyl derivatives of 4 were synthesized (Scheme 2). The desired intermediate chloro derivative (8) was synthesized as shown in Scheme 1. Thus, Toyocamycin 4 was diazotized by a modified procedure [8] to obtain desaminotoyocamycin (6) which was protected as the corresponding triacetate (7) in high yield. Hydroxy derivative 7 was then refluxed [9] in POCl₃ for 16 h to obtain the crucial intermediate 8 in quantitative yield.

The treatment of intermediate 8 with saturated methanolic ammonia at 5 °C for 2.5 h furnished 4-chloro-pyrrolo[2,3-d]pyrimidine nucleoside (9). The derivatives (10–18) modified at C4 position of Toyocamycin 4 were obtained from either of the intermediates 8 or 9. Thus mono N^4 -alkyl derivatives 10–15 (Table 1) were then generated in a small library fashion by exposing 8 to respective amines. Although the larger size of alkyl group has improved the activity more than two fold (10 vs. 13) it also increased the cytotoxicity multiple times. Apart from mono N^4 -alkyl derivatives (10–15), the corresponding dialkyl

Scheme 2. Reagents: (i) Saturated methanolic ammonia; (ii) RNH₂, THF; (iii) NHMe₂, THF (or) NaOMe, MeOH (or) NaSMe, isopropyl alcohol.

Compound	R	$EC_{50}(\mu M)$	$CC_{50}(\mu M)$
10	NHMe	11	300
11	NH—	8	100
12	NH	7	50
13	$NH \sim N \bigcirc O$	4.5	40
14	NH	5	75
15	NH	5	100
16	NMe_2	3	250
17	OCH_3	0.4	40
18	SCH_3	0.4	250

Table 1 SAR of the substitution at C4 position of Toyocamycin

 $(N^4, N^4$ -dimethyl) derivative [9] (16) was also synthesized to understand the influence of substituting both protons of exocyclic amino group at C4 of Toyocamycin 4.

Thus, chloro derivative 9 upon exposure to dimethyl amine in refluxing THF afforded the N^4, N^4 -dimethyl derivative 16. Derivative 16 was found to show better activity than that of mono N^4 -alkyl derivatives 10-15. This indicated that the two protons on the exocyclic amino group at C4 in Toyocamycin 4 were not essential for the activity. This was further validated by the altogether replacement of exocyclic amino group by different hetero-atom groups as in the case of methoxy (17) and methylthio (18) derivatives [9]. These derivatives (17 and 18) were obtained from 9 by the reaction with NaOMe or NaSMe. Between the derivatives 17 and 18, the latter was found to be less toxic than the former. The excellent activity of derivatives 17 and 18 over that of 10-15 might suggest that the hydrogen bond acceptors were better tolerated than the hydrogen bond donors at C4 position in the absence of any modification of cyano group at C5 position of Toyocamycin 4.

After establishing the modifications allowed at C4 position of Toyocamycin 4, to improve the activity of original lead adenosine derivative 1, we then focused our attention on modifications involving both C4 and C5 positions (Table 2). Initially, the C5 cyano group in desaminotoyocamycin 6 was modified. The target derivatives carboxamide

Table 2	
SAR of the substitution at C4 and	C5 positions of Toyocamycin

Compound	$EC_{50}(\mu M)$	$CC_{50}(\mu M)$
19	4	33
20	33	300
21	100	300
22	25	250
23	0.6	200
24	3.7	300
25	9	300
26	2.5	300
27	1	200

(19), carboxamido oxime (20), carboxamidine (21) and thioamide (22) were prepared (Scheme 3) as described previously in literature [8]. Of these 5-carboxamido derivative 19 had shown comparable activity as that of compounds 10–18 derived from single modification. Although carboxamide 19 exhibited good activity but it's also found to be relatively toxic (CC_{50} : 33 μ M), whereas derivatives 20–22 were weakly active but they were not toxic.

This prompted us to explore carboxamide oxime and carboxamidine modifications of cyano group in N^4 -methyl 10 and N^4,N^4 -dimethyl 16 and methylthio 18 derivatives. Accordingly, derivatives 10, 16 and 18 were converted into the corresponding carboxamide oximes (23, 25 and 27), respectively, upon reaction with aqueous hydroxylamine in refluxing ethanol. The corresponding carboxamidines (24 and 26) of the carboxamide oximes 23 and 25 were obtained under catalytic hydrogenation conditions.

Among the carboxamide oximes 20, 23, 25 and 27 of this study only the carboxamide oxime 23 derived from nitrile 10 was found to have highest activity. A closer look of oxime 23 revealed that it also has good selectivity [selectivity index (SI): 333] only next to that of nitrile derivative 18 (SI: 625). Furthermore, the carboxamidines 24 and 26 were also found to have good activity and selectivity over that of 21. This confirms to the fact that absence of hydrogen bond donors, such as substituted amino groups, at C4 of Toyocamycin 4 enhances the activity even when cyano group at C5 is modified. The thioamides (not shown) derived from compounds 10, 16 and 18 were also found to be inactive as that of 22. Thus, simultaneous modification of C4 and C5 positions of Toyocamycin 4 did not enhance the activity to a large extent over modifications at C4 alone.

In conclusion, we have synthesized and screened a variety of Toyocamycin 4 analogues (10–27) for their ability to inhibit HCV RNA replication in replicon cells. The derivatives 17 and 18 from single modifications at C4 and analogues 23 and 27 from combined modifications of C4 and C5 positions of Toyocamycin 4 showed good activity over the rest of the compounds examined in this study. However derivatives 18, 23 and 27 were found to have good selectivity in decreasing order.

Scheme 3. Reagents: (i) H₂O₂, NH₄OH; (ii) aq NH₂OH, EtOH; (iii) H₂S, NEt₃, py.; (iv) Raney Ni, H₂.

References

- [1] (a) B.K. Chun, P. Wang, A. Hassan, J. Du, P.M. Tharnish, L.J. Stuyver, M.J. Otto, R.F. Schinazi, K.A. Watanabe, Tetrahedron Lett. 46 (2005) 2825–2827;
 - (b) J. Shi, J. Du, T. Ma, K.W. Pankiewicz, S.E. Patterson, P.M. Tharnish, T.R. McBrayer, L.J. Stuyver, M.J. Otto, C.K. Chu, Bioorg. Med. Chem. 13 (2005) 1641–1652;
 - (c) E.J. De Clercq, Clin. Virol. 30 (2004) 115-133;
 - (d) R.C. Griffith, L. Lou, C.D. Roberts, U. Schmitz, Annu. Rep. Med. Chem. 39 (2004) 223-237;
 - (e) M.P. Walker, Z. Hong, Curr. Opin. Pharmcol. 2 (2002) 534-540.
- [2] (a) C. Okuse, J.A. Rinaudo, K. Farrar, F. Wells, B.E. Korba, Antiviral Res. 65 (2005) 23–34;
 - (b) Y. Asahina, N. Izumi, N. Enomoto, M. Uchihara, M. Kurosaki, Y. Onuki, Y. Nishimura, K. Ueda, K. Tsuchiya, H. Nakanishi, T. Kitamura, S. Miyake, J. Hepatol. 43 (40) (2005) 623–629.
- [3] (a) K.L. Smith, V.C.H. Lai, B.J. Prigaro, Y. Ding, E. Gunic, J-L. Girardet, W. Zhong, Z. Hong, S. Lang, H. An, Bioorg. Med. Chem. Lett. 14 (2004) 3517–3520;
 - (b) Y. Ding, J.-L. Girardet, K.L. Smith, G. Larson, B. Prigaro, V.C.H. Lai, W. Zhong, J.Z. Wu, Bioorg. Med. Chem. Lett. 15 (2005) 675–678;
 - (c) Y. Ding, H. An, Z. Hong, J.-L. Girardet, Bioorg. Med. Chem. Lett. 15 (3) (2005) 725–727.
- [4] (a) A.R. Porcari, L.B. Townsend, Synth. Commn. 28 (20) (1998) 3835–3843;
 - (b) A.R. Porcari, L.B. Townsend, Nucleosides Nucleotides 18 (1999) 153-159.
- [5] (a) D.E. Bergstrom, A.J. Brattesani, M.K. Ogawa, M.J. Schweickert, J. Org. Chem. 46 (1981) 1423-1431;
 - (b) T. Maruyama, L.L. Wotring, L.B. Townsend, J. Med. Chem. 26 (1983) 25-29;
 - (c) E.D. Clercq, J. Balzarini, D. Madej, F. Hansske, M.J. Robins, J. Med. Chem. 30 (1987) 481-486;
 - (d) S.H. Krawczyk, M.B. Rodriguez, M.R. Nassiri, E.R. Kern, L.L. Wotring, J.C. Drach, L.B. Townsend, J. Med. Chem. 33 (1990) 3160–3169;
 - (e) S.H. Krawczyk, M.R. Nassiri, L.S. Kucera, E.R. Kern, R.G. Ptak, L.L. Wotring, J.C. Drach, L.B. Townsend, J. Med. Chem. 38 (1995) 4106–4114.
- [6] (a) P.K. Gupta, S. Daunert, M.R. Nassiri, L.L. Wotring, J.C. Drach, L.B. Townsend, J. Med. Chem. 32 (1989) 402–408;
 - (b) P.K. Gupta, M.R. Nassiri, L.A. Coleman, L.L. Wotring, J.C. Drach, L.B. Townsend, J. Med. Chem. 32 (1989) 1420–1425;
 - (c) N.K. Saxena, L.A. Coleman, J.C. Drach, L.B. Townsend, J. Med. Chem. 33 (1990) 1980–1983;
 - (d) T.E. Renau, L.L. Wotring, J.C. Drach, L.B. Townsend, J. Med. Chem. 39 (1996) 873-880;
 - (e) T.E. Renau, C. Kennedy, R.G. Ptak, J.M. Breitenbach, J.C. Drach, L.B. Townsend, J. Med. Chem. 39 (1996) 34470–34476.
- [7] J. Shim, G. Larson, V. Lai, S. Naim, J.Z. Wu, Antiviral Res. 58 (2003) 243.
- [8] B.C. Hinshaw, J.F. Gerster, R.K. Robins, L.B. Townsend, J. Org. Chem. 35 (1970) 236–241.
- [9] B.C. Hinshaw, O. Leonoudakis, K.H. Schra, L.B. Townsend, J.C.S. Perkin Trans. I (1975) 1248–1253.